

## Lipid Nanoparticles for Overcoming Biological Barriers to mRNA Delivery to the Brain

Misbaul Hoque<sup>1</sup>, Bidisha Das<sup>2</sup>, Sandra Tiwari<sup>3</sup>, Rupam Roy<sup>4</sup>, Shrestha Sinha Ray<sup>5</sup>, Ankita Bhunia<sup>6</sup>, Soumyarshi Mukhopadhyay<sup>7</sup>, Aishik Bhattacharjee<sup>8</sup>, Proloy Saha<sup>9</sup>, Dr. Logeswar S T<sup>10</sup>, and \*Rounak Bhattacharya

<sup>1</sup>Assistant Professor, Elias Choudhury Academy

<sup>2</sup>Assistant Professor, Dr. Sudhir Chandra Sur Institute of Pharmaceutical Science and Technology.

<sup>3</sup>Department of Pharmacy, Guru Nanak Institute of Pharmaceutical Science & Technology, 157/F Nilgunj Road, Panihati, Sodepur, Kolkata 700114, India

<sup>4</sup>Assistant Professor, Dr. Sudhir Chandra Sur Institute of Pharmaceutical Science and Technology.

<sup>5</sup>Department of Pharmacy, Guru Nanak Institute of Pharmaceutical Science & Technology, 157/F Nilgunj Road, Panihati, Sodepur, Kolkata 700114, India

<sup>6</sup>Department of Pharmacy, Guru Nanak Institute of Pharmaceutical Science & Technology, 157/F Nilgunj Road, Panihati, Sodepur, Kolkata 700114, India

<sup>7</sup>Department of Pharmacy, Guru Nanak Institute of Pharmaceutical Science & Technology, 157/F Nilgunj Road, Panihati, Sodepur, Kolkata 700114, India

<sup>8</sup>Department of Pharmacology, Guru Nanak Institute of Pharmaceutical Science & Technology, 157/F Nilgunj Road, Panihati, Sodepur, Kolkata 700114, India

<sup>9</sup>Department of Pharmacy, Guru Nanak Institute of Pharmaceutical Science & Technology, 157/F Nilgunj Road, Panihati, Sodepur, Kolkata 700114, India

<sup>10</sup>assistant Professor, Department of Pharmacy Practice, JKK Munirajah Institute of Health Sciences College of Pharmacy, T.N.Palayam, Gobi(Tk), Erode (Dt), Tamilnadu-638506

<sup>\*</sup>, Department of Pharmaceutical Chemistry, Guru Nanak Institute of Pharmaceutical Science and Technology, 157/F Nilgunj Road, Panihati, Sodepur, Kolkata 700114, India

Corresponding Email: rounakbhattacharya2002@gmail.com

### Abstract

Background: The advent of messenger RNA (mRNA) as a programmable modality has radically changed the therapeutic environment of neurodegenerative, oncological, and genetic CNS disease. But Blood-Brain Barrier (BBB) and hepatic sequestration are still daunting hurdles to systemic delivery. Purpose: This review is a critical synthesis of the 2019-2026 advances in lipid nanoparticle (LNP) engineering to overcome the neurovascular unit and reach cell-specific tropism. Methods: According to PRISMA-ScR principles, we review peer-reviewed publications dealing with novel ionizable lipid libraries, corona engineering, and alternative routes of administration. Findings: Recent discoveries point to the shift of passive diffusion to active receptor-mediated transcytosis (RMT) by using ligands like SR-57227 (5-HT<sub>3</sub> receptor) and MK16-mediated caveolae pathways. We look at the change in

subcellular mechanics that is the Vesicle Budding-and-Collapse (VBC) model, which states that post-escape aggregate dissolution is a new rate-limiting bottleneck. Clinical feasibility is shown in primate and human *ex vivo* models by therapeutic advances in glioblastoma (IL-12 mRNA) and base editing of congenital diseases (MPS-IH). Conclusions: Although efficiency has improved 50-fold relative to FDA-approved standards (MC3), existing systemic platforms remain able to transfect 1-2 percent of neurons. This review indicates research gaps with regard to sex-based pharmacological differences and the absence of chronic safety profiles. Implications: These intelligent LNPs can offer a strong platform on how to change what used to be fatal neurological conditions into manageable or curable diseases using molecular neuro-restoration.

**Keywords:** Blood-Brain Barrier, Lipid Nanoparticles, mRNA Therapeutics, Endosomal Escape, Neurodegeneration, Receptor-Mediated Transcytosis.

## 1. Introduction:

The introduction of messenger RNA (mRNA) as an adjustable therapeutic modality has completely transformed the landscape of contemporary pharmacology[1]. In the past, genetic, neurodegenerative and oncological diseases of the CNS were impeded due to the difficulty in accessing the large functional biomolecules to the brain. The benefit of mRNA over more conventional protein-based or DNA-based therapies is that it takes advantage of the native translational machinery in the cell to express therapeutic proteins in the cell, and it has no risk of insertional mutagenesis (as seen with viral vectors) and does not need to be taken up into the cell nucleus (as seen with DNA). This flexibility enables mRNA to be an expandable platform to infectious disease vaccines, to targeted neuro-restoration[2].

Even with this potential, the inherent weakness of mRNA is a major challenge; its size (polyanionic) and susceptibility to enzymatic hydrolysis by ribonucleases (RNases) make it necessary to have a complex delivery vehicle. Lipid nanoparticles (LNPs) have emerged as the ideal non-viral delivery system to do this, but there is a significant physiological challenge to systemically administered LNPs: "Liver Tropism". When injected intravenously, typical LNPs, including FDA-approved DLin-MC3-DMA formulation, quickly adsorb a biomolecular corona of host serum proteins[3]. The major constituent of this corona is Apolipoprotein E (ApoE), which assists in clearing of particles to the hepatocytes through the low-density lipoprotein receptor (LDLR) before they can ever enter the cerebral circulation. This is a kind of natural sequestration in the liver, together with the existence of the blood-brain barrier (BBB), which has traditionally rendered the brain an inaccessible target with respect to mRNA-based medicine[4].

The BBB is not a wall but a dynamic, multi-component system, the neurovascular unit (NVU) that combines mechanical, chemical and cellular defenses to sustain neural homeostasis. The endothelial cells which constitute this barrier are microvascular and have no fenestrations and a network of tight junction proteins (such as claudin-5 and occludin) that limit the entry of practically all macromolecules[5]. This has led to unprecedented success of LNPs in liver-targeted therapies and systemic vaccines, but their application to CNS disorders needs to overcome the hepatic sequestration barrier in addition to the inhibitory endothelial interface of the NVU.

The years 2023-2026 have seen a watershed shift in nanomedicine, with the shift in perception of passive carriers to Brain-Targeting Lipid Nanoparticles (BLNPs)[6]. This development marks a radical move towards precision neuro-nanomedicine, with the delivery vehicle as programmable as its genetic payload. Scientists are in the process of preparing smart BLNPs with brain-targeting ligands (ex: serotonin-mimetic OS4T LNP or tryptamine-derived TD5 BLNP), or with corona-engineered surfaces that prevent hepatic uptake. These new-generation platforms take advantage of active transport systems, including receptor-mediated transcytosis (RMT), to cross the BBB. Indicatively, more recent formulations such as the MK16 BLNP have shown mRNA delivery efficiencies 50-fold greater than typical clinical standards, and allowed mRNA delivery to neurons and astrocytes in large brain regions[7]. The development of BLNPs is transforming previously deadly neurological diseases into molecular targets that may be treated by progressing beyond mere encapsulation to active neurovascular navigation.

## 2. Literature Review

This scoping review methodology followed the Preferred Reporting Items for Systematic reviews and Meta-Analyses extension of Scoping Reviews (PRISMA-ScR) to allow a clear and reproducible synthesis of the fast-changing landscape of mRNA-LNP[8]. A systematic literature review was conducted in leading databases covered by the Scopus index, such as PubMed, Web of science, and Google Scholar, with a publication date ranging between January 2019 and April 2026[9]. This particular period was chosen to include the paradigm shift in the delivery system that was previously passive and liver-targeted to the existing generation of the so-called smart brain-targeting lipid

nanoparticles (BLNPs). Keywords were combined with Boolean operators with search query terms including, but not limited to: lipid nanoparticles, ionizable lipids, blood-brain barrier, neurovascular unit, and mRNA-based gene therapy.

Peer-reviewed original studies and high-impact reviews were given preference as inclusion criteria based on mechanistic understanding or high-quality translational data. In particular, only studies with a description of the synthesis of novel ionizable lipid libraries designed to penetrate the CNS, such as serotonin-mimetic (OS4T), tryptamine-derived (TD5), and neurotransmitter-derived (NT1-O12B) formulations were considered[10]. The literature explaining the molecular aspects of barrier crossing was principally kept in focus, particularly active receptor-mediated transcytosis (RMT) utilizing caveolae- and -secretase-mediated pathways as the example of the MK16 platform. The review also focused on studies that use high-throughput *in vivo* screening methods, including barcoded mRNA libraries, that have greatly accelerated the discovery of lipids with high neuronal tropism with reduced hepatic off-target effects[11].

The parameters of critical structural-activity relationship (SAR) were targeted in data extraction, such as optimization of particle pKa (usually 6.2-6.5), functionalization of surfaces with targeting peptides (e.g. RVG29), and the presence of biodegradable ester linkers to enable endosomal escape[12]. The studies focusing on clinical potential using primate and human *ex vivo* models were emphasized to meet 2026 priorities with clinical readiness. Lastly, the selection process put focus on studies that explored biological differences, including sex-related differences in neurovascular integrity and inflammatory response, ensuring the review addresses modern requirements for pharmacological precision and inclusive clinical translation[13].

### **3. The Neurovascular Unit (NVU) as a Dynamic Barrier**

Researchers need to step out of the historic conceptualization of blood-brain barrier (BBB) as a static wall to engineer effective lipid nanoparticles (LNPs) to deliver to the brain. The interface is recognized in modern neurobiology as the neurovascular unit (NVU), a multi-component, highly dynamic system, which combines mechanical, chemical, and cellular defense mechanisms to ensure neural homeostasis[14]. NVU contains brain microvascular endothelial (BMVEC) and pericyte basement membrane embedded pericytes, and astrocyte end-feet processes (glia limitans). This is a collaborative structure that guarantees the brain is guarded against any external toxin and pathogens and that the intake of vital nutrients and extraction of waste materials are precisely controlled.

#### **3.1 Tight Junction Dynamics: Claudin-5 and Occludin.**

The main physical limitation of the NVU lies within the brain microvascular endothelial cells (BMVECs), being completely devoid of fenestrae and having a greatly reduced rate of non-specific transcytosis as opposed to peripheral vasculature[15]. A complicated system of tight junction (TJ) proteins controls the integrity of this paracellular seal. These crossings consist of transmembrane proteins, mainly claudin-5 and occludin and junctional adhesion molecules (JAMs). They are bound to the actin cytoskeleton of the cell through intracellular zonula occlusives (ZO) proteins and form a high-resistance barrier, limiting the free diffusion of even small polar molecules[16]. These specialized junctions are formed by a highly controlled mechanism triggered in fetal life by the Wnt/ $\beta$ -catenin signaling pathway, which causes CNS-specific vascular specialization. This early maturation happens during the first trimester, giving a barrier that is fully functional at the second trimester and creates a considerable barrier to the delivery of large and polyanionic molecules such as mRNA[17].

### **3.2 Blood-CSF Barrier (BCSFB): Paracrine Opportunities**

Although the BBB controls the passage of substances through the blood into the brain parenchyma, the second interface can be found at the choroid plexus: the blood-cerebrospinal fluid barrier (BCSFB)[18]. Despite the fact that the endothelial cells of the choroid plexus are fenestrated, the barrier effect is relocated to the epithelial cells of the choroid plexus which are joined together with tight junctions. New methods of therapy have started to exploit the BCSFB as another route of genetic medicine entry. LNPs may induce the production of therapeutic proteins by acting on the epithelial cells of the choroid plexus and the proteins are then released into the CSF circulation[19]. This paracrine style is especially effective in treating lysosomal storage diseases or diseases that involve the distribution of enzyme throughout the body since the CSF can be used to carry these secreted proteins to periventricular and other cortical regions.

### **3.3 Pathological Changes: Neurodegeneration and Glycocalyx Control**

The NVU is subjected to severe pathological changes that change its permeability and clearance ability in the context of neurodegenerative diseases (NDDs) like Alzheimer (AD), Parkinson (PD), and Multiple Sclerosis (MS). In both AD and PD, the dysfunction of the BBB causes the breakdown of the clearance of neurotoxic proteins, such as beta-amyloid ( $A\beta$ ), tau, and alpha-synuclein, which subsequently build up in the brain parenchyma and speed up neuronal death[20]. This impairment is usually complemented by the mobilization of endothelial cells, which express inflammatory markers and inflammatory cytokines, which leads to a vicious cycle of barrier degradation and neuroinflammation.

Importantly, recent studies point out that a decline of barriers is closely associated with dysregulation of the glycocalyx. A carbohydrate-rich layer of the luminal endothelium is the glycocalyx, which serves as the initial protective barrier[21]. Aging and neurodegeneration Abnormal glycoproteins and a loss of glycocalyx density undermine the selective integrity of the barrier and predispose the brain to micro-hemorrhages and peripheral immune cell infiltration. Moreover, oxidative stress and mitochondrion dysfunction in BMVECs result in the discharge of reactive oxygen species (ROS), which further weakens tight junctions and the work of efflux transporters such as P-glycoprotein (P-gp), making the brain more susceptible to toxic assault[22]. These pathological variations are crucial to 2026 clinical priorities because LNPs need to be designed to function in healthy and diseased neurovascular enclaves.

## **4. Next-Generation LNPs Core Engineering Principles.**

A strict concept of chemical structure-activity relationships (SAR) predicts the transition of lipid nanoparticles (LNPs) as simple systemic delivery vehicles to precision neuro-nanomedicine[23]. Whereas initial LNP development strongly depended on empirical cycles, the 20232026 research environment has shifted to rational, modular paradigms, which adopt molecular precision to deliver to organs. An average LNP is a multi-component structure that is typically 60 to 180 nm in size, and the activity is determined by the exact ratio and chemical structure of four major lipid classes: ionizable, helper phospholipids (e.g. DSPC or DOPE), cholesterol, and PEGylated lipids. Among them, the ionizable lipid is the functional "engine" that not only provides the efficient encapsulation of the polyanionic mRNA cargo but also the subsequent destabilization of endosomal membranes necessary to release the cargo into the cytosol[24].

### **4.1 Ionizable Lipids: pKa Optimization and New Headgroups**

The efficiency of an LNP crucially depends on the pKa of its ionizable lipid that defines the pH at which the molecule changes between a neutral state and cationic one. The current SAR consensus proposes optimal pKa of 6.2 to 6.5 as the sweet spot of in vivo delivery that enables the LNP to stay

near-neutral at physiological pH (7.4) and reduce non-specific interactions and systemic toxicity, and protonate and positively charge at acidic microenvironment (pH < 6.5) of the early endosome[25]. The next generation lipids have gone further than the simple aliphatic amines and have been developed to neurotransmitter-mimetic headgroups that exploit certain pathways in the brain through their receptors. As an example, in the Lipid S4 formulation, a high-affinity ligand of a serotonin [5-hydroxytryptamine type 3 (5-HT<sub>3</sub>)] receptor, SR-57227, is used as the core headgroup. The resultant OS4T LNP has a serotonin-mimetic design, which allows the serotonin-mimetic to increase the brain mRNA expression by 13.3-folds relative to the clinical benchmark, DLin-MC3-dma. Likewise, the TD5 BLNP has a tryptamine-derived headgroup that takes advantage of the endocytosis of 5-HT<sub>1A</sub> receptors to provide excellent central nervous system (CNS) tropism, outcompeting FDA-approved standards by 50-fold in cell cultures[26].

#### **4.2 Structural Motifs: Dual-THF Cores and Biodegradable Linkers**

Recent SAR research has also found that the internal structural motifs of the lipid tails and cores play a major role in determining the stability of the LNP and its capability to travel through the neurovascular unit. Another breakthrough is the creation of the F11T6 LNP with a distinctive dual-tetrahydrofuran (THF) core structure[27]. This bis-THF motif increases stability of lipid bilayers and the strength of interaction with lipoprotein receptors on brain endothelial cells, which leads to neuron-targeted delivery efficiency of 16.4% dose/g- the highest reported so far with systemic CNS-targeting platforms. These next-generation lipids are typically designed to be safe and high-speed cargo accessible, and to achieve this, they may be designed to include biodegradable linkers, e.g., the carbonate ester linkers used in the TD5 formulation or the acetal tails in F11T6. These linkers will undergo hydrolysis upon acid exposure, so that the entire nanoparticle can break down quickly when it reaches the acidic endosome[27]. Not only does this structural transition limit long-term lipid accumulation and neuro-inflammation, but also speeds up intracellular release kinetics, which is essential to ensure that the mRNA cargo is immediately available to the translational apparatus of the target cell.

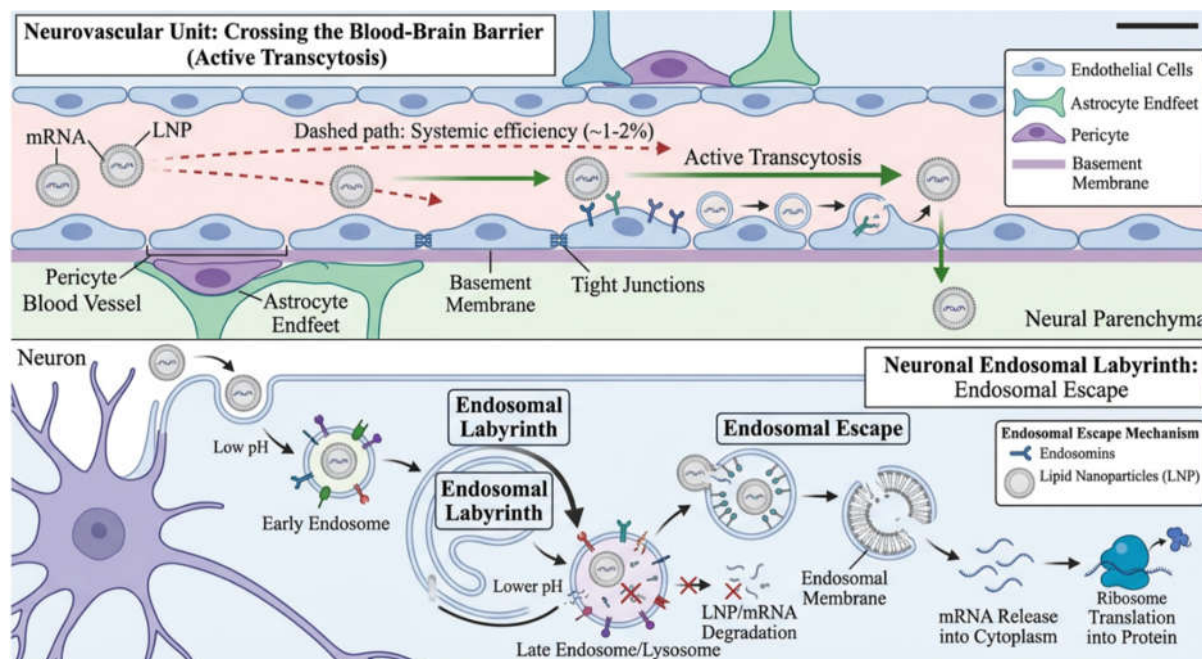
#### **4.3 Corona Engineering: Dead ApoE Strategy of Liver Detargeting**

Although brain-specific ligands have been developed, the liver tropism of systemically delivered LNPs is an uphill challenge. When injected intravenously LNPs adsorb nature biomolecular corona of host serum proteins, which is dominated by Apolipoprotein E (ApoE). This corona enables the swift absorption of LNPs into the hepatocytes through the low-density lipoprotein receptor (LDLR) before the LNPs can enter the brain[28]. To address this, scientists have been the first to engineer corona with dead ApoE (dApoE) mutants. These dApoE variants are designed with targeted replacement in the receptor-binding domain that interrupts the association with the liver LDLR whilst not affecting their capacity to associate to the LNP surface. Coatings of dApoE onto LNPs serve as a molecular mask, which effectively targets the liver and reduces hepatocyte transduction by 90%. This approach does not simply get the particles out of the system but rather prolongs the half-life of the LNPs in circulation, thereby increasing the likelihood of contacts with the cerebral vasculature and allowing brain-targeted ligands to allow receptor-mediated transcytosis to occur over a broader time-span[29]. A dual-functional system can be achieved, using dApoE to target liver and active retargeting to CNS using particular peptide ligands like RVG29.

### **5. Mechanisms of Crossing and Subcellular Escape**

The process of lipid nanoparticles (LNPs) entering the neural parenchyma is a journey that involves a series of steps, where it is necessary to overcome not only the macro-level barrier of the neurovascular unit but also the micro-level labyrinth of the endosomes in the target cells. Although the most scalable route is systemic, its historical efficiency is very poor, only about 1 percent to 2 percent of the dose actually makes it to neurons[30]. To overcome this, 2026 studies have concentrated on the orchestration of active transcytosis precisely and made a fundamental reconsideration of endosomal escape

mechanisms. Overcoming these interlinked biological challenges at a molecular level is now the key to achieving therapeutic relevance in complex CNS disorders.



**Figure 1. LNP Delivery: From Neurovascular Unit to Endosomal Escape**

**5.1 Active Transcytosis: Caveolae/ secretase- and - secretase-mediated Pathways**

One of the breakthroughs in the field of brain delivery was the discovery of the MK16 BLNP platform that exploits active receptor-mediated transcytosis (RMT) to pass through the restrictive endothelial interface without breaking tight junctions[31]. The MK16 platform uses specific cellular machinery to be transported across the endothelial cytoplasm, unlike non-specific adsorption-mediated pathways (AMT), which typically require electrostatic interactions between the particle and anionic cell surfaces. These nanoparticles have been shown to be transported into cells via caveolae-mediated transcytosis, which is a process of membrane invaginations that are small and flask-like. The pathway is critically important as studies have demonstrated that disruption of caveolae formation by depletion of membrane cholesterol, significantly decreases mRNA delivery to the brain[32].

**Table 1. Characteristics and mechanistic components of the MK16 BLNP transcytotic platform. (source: GPAI)**

Feature/Component	Mechanism/Observation	Impact on Brain Delivery
Transport Pathway	Receptor-Mediated Transcytosis (RMT) via Caveolae	Facilitates passage across endothelial interface without disrupting tight junctions.
Cellular Machinery	Caveolae-mediated invaginations) (flask-like)	Dependent on membrane cholesterol; depletion significantly reduces mRNA delivery.

<b>Enzymatic Regulation</b>	-secretase pathway (e.g., inhibited by NGST)	Enables programmed abluminal release into brain parenchyma.
<b>Molecular Mediators</b>	6-hydroxymethyl-4-methoxypyrazine (HMO) complex	Critical component of the transcytotic machinery identified in 2025-2026 studies.
<b>Performance Metric</b>	High-affinity interaction	Surpasses current clinical standards by a factor of >50.

Moreover, the unexpected implication of the MK16 platform is a mechanistic breakthrough, which entails the unexpected role of the proteins of the complex of proteins called 6-hydroxymethyl-4-methoxypyrazine (HMO) in the pathology of Alzheimer disease. Researchers found that in 2025 and 2026, the nanoparticle transcytotic machinery depends on a key component of the transcytotic machinery: the role of the 2025 and 2026 studies demonstrated that the 2025 and 2026 studies were critical components of the transcytotic machinery of a nanoparticle. It was also revealed that pretreatment with the  $\gamma$ -secretase inhibitors, including NGST, can disrupt the delivery of MK16 LNPs to the brain, indicating that LNPs can be chemically programmed to activate a certain set of enzymatic pathways to enable them to abluminal release into the brain parenchyma[33]. Such high-affinity interaction can enable the smart BLNPs to surpass clinical standards by a factor of more than 50.

## 5.2 The Endosomal Escape Revolution: A Change of Model to the VBC

Researchers found that in 2025 and 2026, the nanoparticle transcytotic machinery depends on a key component of the transcytotic machinery: the role of the 2025 and 2026 studies demonstrated that the 2025 and 2026 studies were critical components of the transcytotic machinery of a nanoparticle[34]. It was also revealed that pretreatment with the  $\gamma$ -secretase inhibitors, including NGST, can disrupt the delivery of MK16 LNPs to the brain, indicating that LNPs can be chemically programmed to activate a certain set of enzymatic pathways to enable them to abluminal release into the brain parenchyma. Nevertheless, more recent pharmacological consensus has moved to the Vesicle Budding-and-Collapse (VBC) model. According to this new framework, the ionizable lipids do not physically tear the endosome, but rather cause a topological change of the endosomal membrane by a series of four distinct events: domain formation, budding, collapse and aggregate formation.

In this reaction, the protonated ionizable lipids are attracted to the anionic endosomal membrane forming lipid-enriched domains that then detach to form small vesicles. These vesicles are naturally fragile as a result of the lamellar-non lamellar hexagonal phase transition resulting in an abrupt disintegration, or collapse, freeing the mRNA cargo[35]. The model emphasizes the importance of helper lipids such as DOPE that are specially designed to facilitate such membrane-disruptive phase transitions in the low pH (pH < 6.5) environment of late endosomes. Through these topological changes, over osmotic swelling, the researchers have engineered lipids, including the F11T6 and TD5 series, that release cytosolically far faster and more efficiently than the first-generation formulations.

## 5.3 Cytoplasmic Aggregates: The Secret Cork of Post-Escape Dissolution

Although the endosomal escape occurs successfully, the last obstacle to protein translation is cytoplasmic aggregates. It is shown in the VBC mechanism that the destruction of the endosomal vesicle frequently causes the development of insoluble lipid- and nucleic acid-complexes in the cytosol.

Although the mRNA has technically entered the cytoplasm, it is still trapped in these aggregates and is therefore not available to the ribosomes. This observation can be used to understand a common discrepancy in LNP research: why a large fraction of the cells could display successful endosomal escape signals but a small fraction of cells displays functional protein expression.

The gradual breakup of these aggregates is an undiscovered bottleneck that dictates the scaling of potency of neurotherapeutics. This has led to a new focus in 2026 engineering with the design of lipids containing an acid-triggered hydrolysis and biodegradable linker, e.g. carbonate or acetal tails, to enhance the rapid dissolution of these post-escape aggregates[36]. To translate the genomic potential into therapeutic reality it is crucial to make sure the mRNA cargo is not only released out of the endosome, but also timely "unpacked" out of its lipid shell. The next generation of BLNPs will overcome these subcellular precision challenges and will alter the CNS treatment paradigm to be no longer manageable relief but curative molecular neuro-restoration.

## **6. Diagnostic and Experimental Approaches.**

By substituting empirical testing with high-throughput screening (HTS) and advanced biomimetic modeling, the quick discovery and optimization of brain-targeting lipid nanoparticles (BLNPs) between 2019 and 2026 have been accelerated. Conventional ways of analyzing mRNA delivery, which frequently used one-formulation injections into large cohorts of animals were not enough to manage the chemical variety of new ionizable lipid libraries[37]. One solution to this was the pioneering of in vivo barcoding of LNP libraries; in this method, individual DNA or RNA barcodes are embedded into various LNPs formulations. This enables a complicated combination of dozens or even hundreds of different LNPs to be introduced to one animal. After tissue harvest, deep sequencing (NGS) is applied to determine the number of copies of each barcode in the brain parenchyma, in effect mapping the "potency landscape" of the library in a single experiment. This strategy helped to discover high-performance leads such as the C14-306 LNP that showed a markedly stronger tropism to NeuN<sup>+</sup> neurons and at the same time showed a decreased hepatic off-target expression.

To supplement these in vivo screens are the state-of-the-art models of the BBB-on-a-chip, offering a human-relevant model of mechanistic studies. These microfluidic systems combine the key components of the neurovascular unit (NVU), such as brain microvascular endothelial cells (BMVECs), astrocytes, and pericytes, which are commonly cultured on the basis of human induced pluripotent stem cells (iPSCs)[38]. The models play a vital role in testing the efficiency of receptor-mediated transcytosis (RMT) pathways under a controlled condition that recreates physiological flow and shear stress. As an example, the effective transportation of acetylcholine-conjugated LNPs was recently confirmed with the help of human BBB-on-a-chip models, and they have the capacity to traverse the endothelial layer and transfect underlying neurons. Moreover, they can also monitor barrier integrity in real-time through trans-endothelial electrical resistance (TEER) measurements and high-resolution imaging of paracellular tight junctions.

After an LNP has made it through the neurovascular unit, its capacity to access the cytosol hinges on its capacity to overcome the initial bottleneck: endosomal escape. Researchers have come up with Galectin-based reporter systems to measure this elusive process[39]. The systems take advantage of the biological activity of Galectin proteins (e.g. Gal8 or Gal9), which are naturally present in the cytosol but have a high affinity to glycans on the inner leaflet of endosomal membranes. The glycans are not revealed to the Galectins until the endosomal membrane is damaged by the LNP ionizable lipids. When membranes rupture, a rapid recruitment of Galectin reporters to the damage site elicits a visible signal, which can be activation of green fluorescent protein (GFP) or luciferase[40]. This method gives a clear, real-time measure of endosomal escape efficiency, enabling researchers to determine the types of nanoparticles that are endosomally trapped and released, and those that are successfully translated.

Lastly, the description of these platforms is also backed by a variety of high-resolution imaging methods. The morphology and internal structure of nanoparticles such as OS4T and MK16 are normally confirmed by cryo-transmission electron microscopy (cryo-TEM), and usually have a diameter ranging between 50 to 150 nm[41]. Modern optical methods, like total internal reflection fluorescence (TIRF) and single-molecule FISH (smFISH) imaging, can be used to track single mRNA molecules in the highly crowded cytoplasm of neural cells. Combining these diagnostic and experimental methods, the field has shifted towards Model-Informed Drug Development (MIDD), in which AI-informed predictions and high-throughput experimental measurements converge to augment the likelihood of clinical success of CNS neurotherapeutics[42].

## **7. Therapeutic and Translational Advances**

The clinical potential of brain-targeting lipid nanoparticles (BLNPs) has moved beyond basic proof-of-concept experiments to highly complex so-called molecular neuro-restoration models aimed at treating incurable central nervous system (CNS) diseases[43]. With the shift to the encapsulation of cargo to the engineering of the so-called smart delivery vehicles, researchers can now offer the spatiotemporal regulation of mRNA translation in the neural parenchyma. The features of these advancements are the creation of cell-specific delivery vehicles and the incorporation of stimuli-responsive payloads that reduce systemic toxicity and maximize therapeutic indices.

### **7.1 Neuro-Oncology: Glioblastoma eIL-12 Delivery using OS4T**

Glioblastoma Multiforme (GBM) is among the most aggressive primary brain tumors in part because of its immunosuppressive microenvironment and the limiting nature of blood-tumor barrier (BTB). In 20252026, a breakthrough was made with development of the OS4T LNP, which employed a serotonin [5-hydroxytryptamine type 3 (5-HT<sub>3</sub>)] receptor ligand, SR-57227, and the Tat cell-penetrating peptide to enable active receptor-mediated transcytosis across the BBB[44]. This platform has been used to deliver engineered Interleukin-12 (eIL-12) mRNA, an effective antitumor cytokine, in orthotopic GBM models[45]. Administering trite systemic IL-12 is hampered by severe immune-related adverse events, but eIL-12 construct adds a matrix metalloproteinase (MMP)-sensitive linker that allows activation of the cytokine in the tumor microenvironment where MMPs are highly expressed. The delivery of eIL-12 via OS4T, increased median survival by twofold (37 days vs. 17 days in negative controls) and robustly suppressed tumor growth and induced the expression of M1 phenotype markers by macrophages and microglia. This local immunotherapy is effective in priming the immune system to attack the malignant cells, without causing the systemic inflammatory weight loss that is normally seen in wild-type IL-12.

### **7.2 Genome Editing: Base Editors and CRISPR/Cas9 to Monogenic Diseases.**

The optimization of BLNPs to CRISPR-Cas9 and adenine base editors (ABE) has significantly improved the prospects of developing a one-and-done type of curative therapy, especially in congenital diseases like Mucopolysaccharidosis Type I (MPS-IH). A study involving C3 LNP platform revealed that producing mRNA-based gene editing machinery in utero or in the neonatal period maximizes the dose to body weight and takes advantage of available fetal stem/progenitor cells[46]. C3. By injecting periventricular cells through intracerebroventricular (ICV) injection. MPS LNPs repaired the Idua G -A mutation in mouse models leading to a sustained biochemical repair with IDUA enzyme activity ranging between 7% to 11% of normal in the midbrain and hippocampus. This repair was enough to greatly decrease the build up of toxic glycosaminoglycans (GAGs). Moreover, this platform was proven clinically ready in human ex vivo pediatric brain tissue, with base-editing efficiencies of about 4% observed without apparent off-target genomic damage. At the same time, the TD5 BLNP has become a genome editing leader in the form of intrathecal administration that causes widespread activation of tdTomato in about 30% of neurons and 40% of astrocytes in major brain regions with a safety profile comparable to FDA-approved standards.

### 7.3 Regenerative Medicine: Phytochemical-loaded LNPs for NDDs

Within the context of regenerative medicine, 2026 studies have proposed ionizable pH-responsive lyotropic liquid crystalline nanoassemblies that are designed to treat neurodegenerative diseases (NDDs) using multi-targeted antioxidant and anti-apoptotic mechanisms. These new-generation LNPs, which are usually based on a polyunsaturated lipid (DLin-MC3-DMA) template, entrap complex phytochemical mixtures of quercetin, ginkgolides B and C, and kaempferol to battle the synergistic effect of oxidative stress and neuroinflammation. The mechanistic factors of these lipid nanoassemblies include the activation of the NRF2/ARE antioxidant defense pathway and pro-survival signaling via the mTOR/AKT/BDNF/GSK3 $\beta$  axis[47]. As an example, the addition of quercetin induces mitochondrial biogenesis and stabilizes mitochondrial membrane potential through the scavenging of reactive oxygen species (ROS). These structural nanoassemblies are tailored to go through transition in the acidic endosomal environment to an inverted hexagonal ( $H_{II}$ ) phase, which enables the ability to release drugs accurately intracellularly, and improve the cytoplasmic bioavailability of labile antioxidants. These "biomimetic" platforms, by having the ability to regulate various pathological states in parallel, offer a powerful platform on which to maintain mitochondrial integrity and aiding neurotrophic processes in the aging or diseased CNS. Such a synergistic solution, based on genetic control and direct biochemical neuroprotection, signifies the next stage of the BLNPs as mere delivery systems into fully-fledged therapeutic systems of molecular neuro-restoration.

### 8. Variations, Research Gaps, and Clinical Implications.

To move lipid nanoparticles (LNPs) beyond conceptual models to clinical reality in the central nervous system (CNS), a radical shift in the performance coverage to a strict account of biological and technical constraints is necessary. Although the 2019-2026 era has produced the breakthrough in brain-targeting lipid nanoparticles (BLNPs), the area is challenged with a complicated efficacy-safety paradox. Academic synthesis should be high-impact, and it has to challenge the status quo, and understand the reason why even the most advanced platforms, including MK16, OS4T, and TD5, face significant biological opposition. The need to deal with these differences and holes is not just an additional practice but the driving force behind the realization of the next generation of accuracy neuro-nanomedicine.

#### 8.1 Biological Differences: Sex-Differences and the Gendered NVU.

One of the important, but poorly studied, variables in CNS drug delivery is the effect of biological sex on the structural and functional health of the neurovascular unit (NVU). Even though present-day research protocols, including the ones that test the OS4T LNP, seem to incorporate both men and women in their studies, the area does not have a fine-grained insight into how hormonal changes and sexually dimorphic gene-expressions patterns determine LNP transcytosis efficiency[48]. More recent academic interest has centered on understanding whether changes in tight junction density (regulated by proteins such as claudin-5 and occludin) or differences in the baseline inflammatory state of the brain determine nanoparticle passage.

Indicatively, more autoimmune neuroinflammation in females indicates a more reactive NVU that can potentially support more adsorption-mediated transcytosis and at the same time, increase the chances of immunotoxic side effects[49]. On the other hand, testosterone and estrogen are also known to increase or decrease the expression of efflux transporters such as P-glycoprotein (P-gp) and BCRP which are capable of actively expelling the synthetic lipids in the brain parenchyma. The current one-size-fits-all dosing regimen applied in mouse models does not take into consideration these physiological peculiarities. Sex-inclusive pharmacokinetics should be one of the priorities of future clinical translation because a delivery system that works in a male model to reach therapeutic concentrations may be sub-therapeutic or toxic in a female model because of unique receptor-mediated transcytosis (RMT) interactions.

## 8.2 Developmental Differences: Maturation of the Barrier.

Another axis of critical variation is developmental stage. The blood-brain barrier (BBB) is not an inert object it is a developing system. The BBB in humans starts to specialize at the fetal stage through the Wnt/  $\beta$ -catenin signalling pathway, and adult tight junction proteins are formed by the second trimester[50]. This premature development is a challenge and an opportunity. A study with the C3 LNP platform has shown that in utero injection can result in a 17-fold enhancement of mRNA expression over neonatal models, primarily due to the availability of plentiful and highly proliferative stem and progenitor cells in the fetal environment.

Nevertheless, when the barrier reaches adulthood the paracellular pathways become narrower and narrower and the non-specific transcytosis rate of the barrier plummets. This development will also require the replacement of the relatively simple delivery strategies that work in the neonate to the highly active, ligand-dependent mechanisms that are needed in the adult brain[51]. The knowledge of these transitions is crucial in the management of monogenic congenital diseases, in which there is a risk of the window of intervention closing as the NVU attains its restrictive adult form.

## 8.3 Research Gaps: The 1% Efficiency Hurdle and Scaling Realities.

Although the next-generation BLNPs are architecturally brilliant, there is a grim research gap: the 1% Efficiency hurdle. In the brain, current systemic vectors, such as those that act on neurons with RVG29 or serotonin-mimetic ligands, usually just transfect 1% to 2% of the total neurons[52]. This can be adequate to depot-style enzyme replacement therapies, in which periventricular cells release enzymes into the CSF to diffuse throughout the brain, but it is severely insufficient in the case of diffuse neurodegenerative diseases, such as ALS or Alzheimer.

To scale this efficiency, it is not sufficient to simply raise the dose but it is necessary to have a fundamental redesign of LNP-cell interactions. Raising dose to reach 10% of neurons would probably lead to hepatic and spleen toxicity, since even brain-targeted particles will be sequestered in large quantities in these clearance organs[53]. The difference is that we cannot reach absolute organ specificity. Indicatively, even the F11T6 LNP, with a high neuron-targeted efficiency of 16.4% dose/g, has significant liver accumulation. To close this gap, 2026 researchers should shift towards the use of multi-stage targeting methods which take advantage of corona engineering (e.g., "dead" ApoE mutants) to target the liver without, at the same time, using high-affinity CNS ligands.

## 8.4 Subcellular Precision: The Invisible Bottleneck

Functional protein translation is inconsistent even in the case an LNP manages to cross the BBB and enter a neuron. This points to a research gap of critical importance of subcellular precision. Quantitative studies show that while 3% to 7% of internalized mRNA might technically "escape" the endosome, much of it remains sequestered in insoluble cytoplasmic aggregates of lipids and nucleic acids[54]. This is one of the insights of the Vesicle Budding-and-Collapse (VBC) model, which demonstrates that escape into the endosome is merely half of the intracellular battle.

The inability of the mRNA to get to the ribosomes regardless of having successfully escaped the membranes is a hidden rate-limiting step. Currently the focus is on the creation of so-called disintegratable lipids, which have acid-dependent hydrolysis or biodegradable carbonate linkers that not only facilitate escape, but also the release of these post-escape aggregates[55]. Unless the kinetics of unpacking in the cytoplasm are addressed, mRNA therapeutics still will be plagued by low potency, no matter how effectively they can bypass the BBB.

### **8.5 Clinical Implications: Long-Term Immunotoxicity and Repeated Dosing**

With the shift of BLNPs to the paradigms of chronic treatment of neurodegeneration, the clinical importance of repeated administration of synthetic lipids should be considered. The immune environment of the brain is highly delicate to foreign materials. Although single dose studies of lipids such as TD5 demonstrate little acute inflammation against clinical standards such as MC3, the impact of decadal dosing is yet to be established. The possibility of a chronic build-up of unmetabolised lipids as microglial and astrocyte lipidosis could provoke a slow-smouldering neuroinflammatory reaction that neutralises the therapeutic effects of the mRNA cargo[56].

Moreover, there is a serious clinical risk of the so-called PEG dilemma. PEGylated lipids are necessary to achieve stability and circulation, but repeated exposure may cause the formation of anti-PEG antibodies leading to the rapid clearance of subsequent doses and possibly severe hypersensitivity. In 2026, clinical readiness will require a shift toward bio-orthogonal or stealthy coating that

### **9. Limitations and Future Directions**

Notwithstanding the fast development of brain-targeting lipid nanoparticles (BLNPs), there are still considerable challenges that require moving toward screening to computational accuracy[57]. Today, functional mRNA transfection of total neurons in the brain is generally only accomplished by systemic administration in 1-2% of the total population, which is usually not enough to treat diffuse neurodegenerative diseases like Alzheimer's or ALS. Moreover, the liver tropism of typical formulations has been found to be persistent; dosage escalation to enhance neural uptake is life-threatening in catastrophic hepatotoxicity as a result of natural ApoE-mediated sequestration in clearing organs[58]. The future of breaking these limitations is in integrating the Artificial Intelligence (AI) and Model-Informed Drug Development (MIDD). AI-guided paradigms are currently applied to forecast the blood-brain barrier (BBB) permeability of multifaceted synthetic lipid libraries and targeting ligands, which essentially maps the "potency landscape" and speeds up the process of identifying high-performance candidates without the time-intensive and costly exhaustive nature of the conventional iterative in vivo screening.

At the same time, MIDD is a foundation of clinical preparedness, which mimics human-specific pharmacokinetics and pharmacodynamics[59]. This is especially important as preclinical rodent models do not necessarily keep the structural complexity and tight junction dynamics of the human neurovascular unit. With MIDD, researchers can use dose selection to optimize the human-relevant dose response profile so that they can keep the therapeutic window and reduce the possibility of chronic immunotoxicity or a type of synthetic lipidosis caused by chronic administration. Further directions should also capitalize on these models to overcome the subcellular precision challenge, or designing lipids that do not only promote endosomal escape by the Vesicle Budding-and-Collapse (VBC) model, but also that dissolve post-escape cytoplasmic aggregates rapidly[60]. Finally, there will be a meeting point between AI-mediated molecular design and computational modeling to bridge the gap between proof-of-concept murine experiments and achievement of safe, high-potency molecular neuro-restoration in humans.

### **10. Conclusion**

In short, the shift between mouse-to-human therapies, as successful and safe and effective, cannot be implemented without a stringent challenge with biological differences and systemic defects. High-impact synthesis should highlight the point that a 1% transfection rate is a starting point and not a limit and that absolute organ specificity by corona engineering is the only way to avoid hepatic toxicity. The dimorphic reactions of the NVU in sex and age and the subcellular puzzle of unpackaging can at last be resolved and the field of BLNPs can be transformed into the life of patients with untreatable CNS conditions. It is evident in the 2026 requirement: we need to design to the complexity of human brain, not the limits of the blood-brain barrier.

The development of smart brain-targeting lipid nanoparticles (BLNPs) is a breakthrough in nanomedicine, and mRNA therapeutics are no longer passively delivered by the system but a programmable neuro-restoration system. Using active transport pathways such as caveolae-mediated transcytosis and Gamma-secretase dynamics, platforms such as MK16 and OS4T have circumvented the blood-brain barrier by 50-fold more efficiently than clinical standards. Despite the major obstacles to the scaling of potency and long-term safety, the combination of AI-directed design and the Vesicle Budding-and-Collapse model offers a strong platform to apply to clinical translation. Finally, BLNPs provide a clear roadmap towards converting incurable or untreatable neurological diseases into treatable or controllable molecular targets.

**Reference:**

- [1] S. N. Heendeniya *et al.*, “Beginning of a new era of synthetic messenger RNA therapeutics: Comprehensive insights on mRNA drug design, development and applications.,” *Exp. Biol. Med. (Maywood)*, vol. 250, p. 10784, 2025, doi: 10.3389/ebm.2025.10784.
- [2] O. Chabanovska, A. M. Galow, R. David, and H. Lemcke, “mRNA – A game changer in regenerative medicine, cell-based therapy and reprogramming strategies,” Dec. 2021, *Elsevier B.V.* doi: 10.1016/j.addr.2021.114002.
- [3] K. Swetha *et al.*, “Recent Advances in the Lipid Nanoparticle-Mediated Delivery of mRNA Vaccines,” Mar. 2023, *MDPI*. doi: 10.3390/vaccines11030658.
- [4] V. I. Teichberg, “From the liver to the brain across the blood-brain barrier,” May 2007. doi: 10.1073/pnas.0702450104.
- [5] C. Greene, N. Hanley, and M. Campbell, “Claudin-5: Gatekeeper of neurological function,” Jan. 2019, *BioMed Central Ltd.* doi: 10.1186/s12987-019-0123-z.
- [6] T. Ilić *et al.*, “Parenteral Lipid-Based Nanoparticles for CNS Disorders: Integrating Various Facets of Preclinical Evaluation towards More Effective Clinical Translation,” Feb. 2023, *MDPI*. doi: 10.3390/pharmaceutics15020443.
- [7] C. Wang *et al.*, “Blood–brain-barrier-crossing lipid nanoparticles for mRNA delivery to the central nervous system,” *Nat. Mater.*, vol. 24, pp. 1653–1663, Oct. 2025, doi: 10.1038/s41563-024-02114-5.
- [8] A. C. Tricco *et al.*, “PRISMA extension for scoping reviews (PRISMA-ScR): Checklist and explanation,” Oct. 2018, *American College of Physicians*. doi: 10.7326/M18-0850.
- [9] M. D. J. Peters *et al.*, “Scoping reviews: reinforcing and advancing the methodology and application,” Dec. 2021, *BioMed Central Ltd.* doi: 10.1186/s13643-021-01821-3.
- [10] F. Ma *et al.*, “Neurotransmitter-derived lipidoids (NT-lipidoids) for enhanced brain delivery through intravenous injection,” *Sci. Adv.*, vol. 6, Jul. 2020, doi: 10.1126/sciadv.abb4429.
- [11] A. G. Hamilton *et al.*, “High-Throughput In Vivo Screening Using Barcoded mRNA Identifies Lipid Nanoparticles With Extrahepatic Tropism for In Situ Immunoengineering,” *Advanced Materials*, Mar. 2026, doi: 10.1002/adma.202514370.
- [12] M. Dastpeyman *et al.*, “Endosomal escape cell-penetrating peptides significantly enhance pharmacological effectiveness and CNS activity of systemically administered antisense oligonucleotides,” *Int. J. Pharm.*, vol. 599, Apr. 2021, doi: 10.1016/j.ijpharm.2021.120398.
- [13] P. Honarpisheh and L. D. McCullough, “Sex as a biological variable in the pathology and pharmacology of neurodegenerative and neurovascular diseases,” Nov. 2019, *John Wiley and Sons Inc.* doi: 10.1111/bph.14675.
- [14] E. C. Kugler, J. Greenwood, and R. B. MacDonald, “The ‘Neuro-Glial-Vascular’ Unit: The Role of Glia in Neurovascular Unit Formation and Dysfunction,” Sep. 2021, *Frontiers Media S.A.* doi: 10.3389/fcell.2021.732820.
- [15] H. L. McConnell, C. N. Kersch, R. L. Woltjer, and E. A. Neuwelt, “The translational significance of the neurovascular unit,” Jan. 2017, *American Society for Biochemistry and Molecular Biology Inc.* doi: 10.1074/jbc.R116.760215.

- [16] L. S. Rodgers and A. S. Fanning, "Regulation of epithelial permeability by the actin cytoskeleton," Dec. 2011. doi: 10.1002/cm.20547.
- [17] M. Corada *et al.*, "The Wnt/ $\beta$ -catenin pathway modulates vascular remodeling and specification by upregulating Dll4/notch signaling," *Dev. Cell*, vol. 18, pp. 938–949, Mar. 2010, doi: 10.1016/j.devcel.2010.05.006.
- [18] S. R. Archie, A. Al Shoyaib, and L. Cucullo, "Blood-brain barrier dysfunction in CNS disorders and putative therapeutic targets: An overview," Nov. 2021, *MDPI*. doi: 10.3390/pharmaceutics13111779.
- [19] A. K. Madadi and M. J. Sohn, "Advances in Intrathecal Nanoparticle Delivery: Targeting the Blood–Cerebrospinal Fluid Barrier for Enhanced CNS Drug Delivery," Aug. 2024, *Multidisciplinary Digital Publishing Institute (MDPI)*. doi: 10.3390/ph17081070.
- [20] X. Yu, C. Ji, and A. Shao, "Neurovascular Unit Dysfunction and Neurodegenerative Disorders," Apr. 2020, *Frontiers Media S.A.* doi: 10.3389/fnins.2020.00334.
- [21] S. M. Shi *et al.*, "Glycocalyx dysregulation impairs blood–brain barrier in ageing and disease," *Nature*, vol. 639, pp. 985–994, Mar. 2025, doi: 10.1038/s41586-025-08589-9.
- [22] N. S. Togra, P. S. Bhoj, N. Mekala, R. Hancock, J. Trivedi, and Y. Persidsky, "Purinergic and extracellular vesicle signaling in alcohol-induced blood–brain barrier breakdown and neuroimmune activation," Nov. 2025, *Academic Press Inc.* doi: 10.1016/j.bbi.2025.106115.
- [23] W. Wang *et al.*, "Artificial intelligence-driven rational design of ionizable lipids for mRNA delivery," *Nature Communications*, vol. 15, Dec. 2024, doi: 10.1038/s41467-024-55072-6.
- [24] P. Mangla, Q. Vicentini, and A. Biscans, "Therapeutic Oligonucleotides: An Outlook on Chemical Strategies to Improve Endosomal Trafficking," Sep. 2023, *Multidisciplinary Digital Publishing Institute (MDPI)*. doi: 10.3390/cells12182253.
- [25] M. J. Carrasco *et al.*, "Ionization and structural properties of mRNA lipid nanoparticles influence expression in intramuscular and intravascular administration," *Commun. Biol.*, vol. 4, Dec. 2021, doi: 10.1038/s42003-021-02441-2.
- [26] D. Cao *et al.*, "Lipid nanoparticles for mRNA delivery in brain via systemic administration," *Science Advances*, vol. 11, Aug. 2025, doi: 10.1126/sciadv.adw0730.
- [27] J. Wang *et al.*, "Recent Advances in Lipid Nanoparticles and Their Safety Concerns for mRNA Delivery," Oct. 2024, *Multidisciplinary Digital Publishing Institute (MDPI)*. doi: 10.3390/vaccines12101148.
- [28] Y. Qi *et al.*, "Flower-Shaped Lipid Nanoparticles Evade Apolipoprotein E-Mediated Liver Tropism for Safe and Enhanced Cytokine-Based Cancer Immunotherapy," *ACS Nano*, vol. 19, pp. 35675–35691, Oct. 2025, doi: 10.1021/acsnano.5c11552.
- [29] Y. Xinchun, T. Jing, and G. Jiaoqiong, "Lipid-based nanoparticles via nose-to-brain delivery: a mini review," *Front. Cell Dev. Biol.*, vol. 11, p. 1214450, 2023, doi: 10.3389/fcell.2023.1214450.
- [30] P. Khare, S. X. Edgecomb, C. M. Hamadani, E. E. L. Tanner, and D. S. Manickam, "Lipid nanoparticle-mediated drug delivery to the brain," Jun. 2023, *Elsevier B.V.* doi: 10.1016/j.addr.2023.114861.

- [31] A. S. Haqqani, K. Bélanger, and D. B. Stanimirovic, "Receptor-mediated transcytosis for brain delivery of therapeutics: receptor classes and criteria," 2024, *Frontiers Media SA*. doi: 10.3389/fddev.2024.1360302.
- [32] J. Sohn, H. Lin, M. R. Fritch, and R. S. Tuan, "Influence of cholesterol/caveolin-1/caveolae homeostasis on membrane properties and substrate adhesion characteristics of adult human mesenchymal stem cells," *Stem Cell Res. Ther.*, vol. 9, Apr. 2018, doi: 10.1186/s13287-018-0830-4.
- [33] C. Wang *et al.*, "Blood-brain-barrier-crossing lipid nanoparticles for mRNA delivery to the central nervous system," *Nat. Mater.*, vol. 24, pp. 1653–1663, Oct. 2025, doi: 10.1038/s41563-024-02114-5.
- [34] X. Liu *et al.*, "Transcytotic transportation of size-controlled nanocarriers into dystrophic skeletal muscle leads to therapeutic outcome in mice," *Nature Communications*, vol. 16, Dec. 2025, doi: 10.1038/s41467-025-66061-8.
- [35] L. Hagedorn, D. C. Jürgens, O. M. Merkel, and B. Winkeljann, "Endosomal escape mechanisms of extracellular vesicle-based drug carriers: lessons for lipid nanoparticle design," Jul. 2024, *OAE Publishing Inc*. doi: 10.20517/evcna.2024.19.
- [36] T. Li *et al.*, "Engineered internal architecture of core-shell lipid nanoparticles promotes efficient mRNA endosomal release," *Nature Communications*, vol. 17, Dec. 2026, doi: 10.1038/s41467-026-69017-8.
- [37] X. Hou, T. Zaks, R. Langer, and Y. Dong, "Lipid nanoparticles for mRNA delivery," Dec. 2021, *Nature Research*. doi: 10.1038/s41578-021-00358-0.
- [38] D. J. Alcendor *et al.*, "Neurovascular unit on a chip: Implications for translational applications," Dec. 2013. doi: 10.1186/srct379.
- [39] S. Chatterjee, E. Kon, P. Sharma, and D. Peer, "Endosomal escape: A bottleneck for LNP-mediated therapeutics," *Proc. Natl. Acad. Sci. U. S. A.*, vol. 121, Mar. 2024, doi: 10.1073/pnas.2307800120.
- [40] S. Omo-Lamai *et al.*, "Limiting endosomal damage sensing reduces inflammation triggered by lipid nanoparticle endosomal escape," *Nat. Nanotechnol.*, vol. 20, pp. 1285–1297, Sep. 2025, doi: 10.1038/s41565-025-01974-5.
- [41] R. Willson, C. Potter, B. Carragher, and A. M. Mulder, "Cryo Transmission Electron Microscopy as an Orthogonal Tool for Characterization of Biopharmaceuticals," <https://pmc.ncbi.nlm.nih.gov/articles/PMC3630664/>, 2012.
- [42] K. Raman, R. Kumar, C. J. Musante, and S. Madhavan, "Integrating Model-Informed Drug Development With AI: A Synergistic Approach to Accelerating Pharmaceutical Innovation," Jan. 2025, *John Wiley and Sons Inc*. doi: 10.1111/cts.70124.
- [43] Y. Wu and A. Angelova, "Recent Uses of Lipid Nanoparticles, Cell-Penetrating and Bioactive Peptides for the Development of Brain-Targeted Nanomedicines against Neurodegenerative Disorders," Dec. 2023, *Multidisciplinary Digital Publishing Institute (MDPI)*. doi: 10.3390/nano13233004.
- [44] K. Wang, "Innovative nanoparticle-based therapeutic strategies against glioblastoma multiform: a focus on enhanced delivery systems and efficacy," 2025, *Frontiers Media SA*. doi: 10.3389/fbioe.2025.1601673.

- [45] S. Roy, D. Alday, and Q. Cai, “Advances in PLGA-Based Drug Delivery Systems for Glioblastoma Treatment,” 2025, *Dove Medical Press Ltd.* doi: 10.2147/IJN.S563730.
- [46] R. Palanki *et al.*, “Ionizable Lipid Nanoparticles for Therapeutic Base Editing of Congenital Brain Disease,” *ACS Nano*, vol. 17, pp. 13594–13610, Jul. 2023, doi: 10.1021/acsnano.3c02268.
- [47] T. Akanchise, F. Luo, B. Angelov, Y. Deng, and A. Angelova, “Designed Liquid Crystalline Nanoassemblies From Clinically Validated Polyunsaturated Lipids for Combined Antioxidant, Anti-Apoptotic, and Neurotrophic Treatments,” *Adv. Healthc. Mater.*, 2026, doi: 10.1002/adhm.202505595.
- [48] M. Simard, P. R. Provost, and Y. Tremblay, “Sexually dimorphic gene expression that overlaps maturation of type II pneumocytes in fetal mouse lungs,” *Reproductive Biology and Endocrinology*, vol. 4, May 2006, doi: 10.1186/1477-7827-4-25.
- [49] G. Marano *et al.*, “Neuroinflammation and the Female Brain: Sex-Specific Mechanisms Underlying Mood Disorders and Stress Vulnerability,” *Life*, vol. 16, p. 139, Jan. 2026, doi: 10.3390/life16010139.
- [50] S. Logan *et al.*, “Studying Human Neurological Disorders Using Induced Pluripotent Stem Cells: From 2D Monolayer to 3D Organoid and Blood Brain Barrier Models,” *Compr. Physiol.*, vol. 9, pp. 565–611, Apr. 2019, doi: 10.1002/j.2040-4603.2019.tb00076.x.
- [51] B. Pawar *et al.*, “Current Update on Transcellular Brain Drug Delivery,” Dec. 2022, *MDPI*. doi: 10.3390/pharmaceutics14122719.
- [52] W. M. Pardridge, “Brain gene therapy with Trojan horse lipid nanoparticles,” May 2023, *Elsevier Ltd.* doi: 10.1016/j.molmed.2023.02.004.
- [53] D. Chenthamara *et al.*, “Therapeutic efficacy of nanoparticles and routes of administration,” Nov. 2019, *BioMed Central Ltd.* doi: 10.1186/s40824-019-0166-x.
- [54] M. Maugeri *et al.*, “Linkage between endosomal escape of LNP-mRNA and loading into EVs for transport to other cells,” *Nat. Commun.*, vol. 10, Dec. 2019, doi: 10.1038/s41467-019-12275-6.
- [55] S. Zhao *et al.*, “Acid-degradable lipid nanoparticles enhance the delivery of mRNA,” *Nat. Nanotechnol.*, vol. 19, pp. 1702–1711, Nov. 2024, doi: 10.1038/s41565-024-01765-4.
- [56] J. Marschallinger *et al.*, “Lipid-droplet-accumulating microglia represent a dysfunctional and proinflammatory state in the aging brain,” *Nat. Neurosci.*, vol. 23, pp. 194–208, Feb. 2020, doi: 10.1038/s41593-019-0566-1.
- [57] M. Sela *et al.*, “AI-Validated Brain Targeted mRNA Lipid Nanoparticles with Neuronal Tropism,” *ACS Nano*, vol. 19, pp. 36106–36128, Oct. 2025, doi: 10.1021/acsnano.4c15013.
- [58] A. Sarode *et al.*, “Potent Liver-Tropic mRNA Lipid Nanoparticles: ApoE-Mediated Delivery Through a Low-Density Lipoprotein Receptor Independent Uptake Mechanism,” *Advanced Materials*, 2025, doi: 10.1002/adma.202517893.
- [59] C. R. Rayner *et al.*, “Model-Informed Drug Development for Anti-Infectives: State of the Art and Future,” Apr. 2021, *Nature Publishing Group*. doi: 10.1002/cpt.2198.
- [60] D. Pei, “Endosomal Escape of Lipid Nanoparticles: A Perspective on the Literature Data,” Dec. 2025, *American Chemical Society*. doi: 10.1021/acsnano.5c11721.